## => d 18 1-11 ti py au so

- L8 ANSWER 1 OF 11 MEDLINE on STN
- TI Detection of beta 2-adrenergic receptor dimerization in living cells using bioluminescence resonance energy transfer (BRET).
- PY 2000
- AU Angers S; Salahpour A; Joly E; Hilairet S; Chelsky D; Dennis M; Bouvier M
- Proceedings of the National Academy of Sciences of the United States of America, (2000 Mar 28) 97 (7) 3684-9.

  Journal code: 7505876. ISSN: 0027-8424.
- L8 ANSWER 2 OF 11 MEDLINE on STN
- TI Role of endocytosis in the activation of the extracellular signal-regulated kinase cascade by sequestering and nonsequestering G protein-coupled receptors.
- PY 2000
- AU Pierce K L; Maudsley S; Daaka Y; Luttrell L M; Lefkowitz R J
- SO Proceedings of the National Academy of Sciences of the United States of America, (2000 Feb 15) 97 (4) 1489-94.

  Journal code: 7505876. ISSN: 0027-8424.
- L8 ANSWER 3 OF 11 MEDLINE on STN
- TI The beta2-adrenergic receptor/betaarrestin complex recruits the clathrin adaptor AP-2 during endocytosis.
- PY 1999
- AU Laporte S A; Oakley R H; Zhang J; Holt J A; Ferguson S S; Caron M G; Barak L S
- SO Proceedings of the National Academy of Sciences of the United States of America, (1999 Mar 30) 96 (7) 3712-7.

  Journal code: 7505876. ISSN: 0027-8424.
- L8 ANSWER 4 OF 11 MEDLINE on STN
- TI Agonist-induced endocytosis and recycling of the gonadotropin-releasing hormone receptor: effect of beta-arrestin on internalization kinetics.
- PY 1998
- AU Vrecl M; Anderson L; Hanyaloglu A; McGregor A M; Groarke A D; Milligan G; Taylor P L; Eidne K A
- SO Molecular endocrinology (Baltimore, Md.), (1998 Dec) 12 (12) 1818-29.

  Journal code: 8801431. ISSN: 0888-8809.
- L8 ANSWER 5 OF 11 MEDLINE on STN
- TI Molecular mechanisms of G protein-coupled receptor desensitization and resensitization.
- PY 1998
- AU Ferguson S S; Zhang J; Barak L S; Caron M G
- SO Life sciences, (1998) 62 (17-18) 1561-5. Ref: 22 Journal code: 0375521. ISSN: 0024-3205.
- L8 ANSWER 6 OF 11 CAPLUS COPYRIGHT 2004 ACS on STN
- TI Arrestin isoforms dictate differential kinetics of A2B adenosine receptor trafficking
- PY 2000
- AU Mundell, Stuart J.; Matharu, Anne-Lise; Kelly, Eamonn; Benovic, Jeffrey L.
- SO Biochemistry (2000), 39(42), 12828-12836 CODEN: BICHAW; ISSN: 0006-2960
- L8 ANSWER 7 OF 11 CAPLUS COPYRIGHT 2004 ACS on STN
- TI Selective regulation of endogenous G protein-coupled receptors by arrestins in HEK293 cells
- PY 2000
- AU Mundell, Stuart J.; Benovic, Jeffrey L.

- SO Journal of Biological Chemistry (2000), 275(17), 12900-12908 CODEN: JBCHA3; ISSN: 0021-9258
- L8 ANSWER 8 OF 11 CAPLUS COPYRIGHT 2004 ACS on STN
- TI Detection of  $\beta$ 2-adrenergic receptor dimerization in living cells using bioluminescence resonance energy transfer (BRET)
- PY 2000
- AU Angers, Stephane; Salahpour, Ali; Joly, Eric; Hilairet, Sandrine; Chelsky, Dan; Dennis, Michael; Bouvier, Michael
- SO Proceedings of the National Academy of Sciences of the United States of America (2000), 97(7), 3684-3689 CODEN: PNASA6; ISSN: 0027-8424
- L8 ANSWER 9 OF 11 CAPLUS COPYRIGHT 2004 ACS on STN
- TI Role of endocytosis in the activation of the extracellular signal-regulated kinase cascade by sequestering and nonsequestering G protein-coupled receptors
- PY 2000
- AU Pierce, Kristen L.; Maudsley, Stuart; Daaka, Yehia; Luttrell, Louis M.; Lefkowitz, Robert J.
- SO Proceedings of the National Academy of Sciences of the United States of America (2000), 97(4), 1489-1494 CODEN: PNASA6; ISSN: 0027-8424
- L8 ANSWER 10 OF 11 CAPLUS COPYRIGHT 2004 ACS on STN
- TI Agonist-induced endocytosis and recycling of the gonadotropin-releasing hormone receptor: Effect of  $\beta\text{-}$  arrestin on internalization kinetics
- PY 1998
- AU Vrecl, Milka; Anderson, Lorraine; Hanyaloglu, Aylin; McGregor, Alison M.; Groarke, Alex D.; Milligan, Graeme; Taylor, Philip L.; Eidne, Karin A.
- SO Molecular Endocrinology (1998), 12(12), 1818-1829 CODEN: MOENEN; ISSN: 0888-8809
- L8 ANSWER 11 OF 11 CAPLUS COPYRIGHT 2004 ACS on STN
- TI Molecular mechanisms of G protein-coupled receptor desensitization and resensitization
- PY 1998
- AU Ferguson, Stephen S. G.; Zhang, Jie; Barak, Larry S.; Caron, Marc G.
- SO Life Sciences (1998), 62(17/18), 1561-1565 CODEN: LIFSAK; ISSN: 0024-3205
- => d his

(FILE 'HOME' ENTERED AT 16:44:05 ON 16 AUG 2004)

- FILE 'MEDLINE, CAPLUS' ENTERED AT 16:44:24 ON 16 AUG 2004
- L1 144 S ?ARRESTIN AND GREEN FLUORESCENT PROTEIN
- L2 46 S L1 AND GPCR
- L3 26 S L2 AND PY <2001
- L4 524 S ?ARRESTIN AND INTERNALIZATION
- L5 380 S L4 AND AGONIST
- L6 102 S L5 AND GPCR
- L7 41 S L6 AND PY <2001
- L8 11 S L7 (L) L3

	Туре	L#	Hits	Search Text	DBs	Time Stamp	Comment s
1	BRS	L1	0	gpcr adj arrestin	USPA T	2004/08/1 6 14:40	
2	BRS	L2	51860	gpcr pathway	1	2004/08/1 6 14:40	
3	BRS	L3	38	12 and arrestin		2004/08/1 6 14:50	
4	BRS	L4	5	13 and @py <2001	USPA T	2004/08/1 6 14:41	
5	BRS	L5	17890	12 and detection	USPA T	2004/08/1 6 14:51	
6	BRS	L6	1223	12 and gfp	•	2004/08/1 6 14:51	
7	BRS	L7	14	13 and gfp	:	2004/08/1 6 14:51	

	U	1	PT	P	Document ID	Issue Date	Pages	Title
1					US 6159707 A	20001212	61	Sperm receptors
2	×				US 6110693 A	20000829	21	Methods of assaying receptor activity and constructs useful in such methods
3					US 5891646 A	19990406	23	Methods of assaying receptor activity and constructs useful in such methods
4					US 5591618 A	19970107	46	G protein-coupled receptor kinase GRK6
5					US 5532151 A	19960702	48	G protein-coupled receptor kinase GRK6

	Current OR	Current XRef	Retrieval Classif	Inventor	s	С	2	3	4	5
1	435/69.1	435/252.3; 435/254.11; ; 435/320.1; 435/325; 435/7.1; 530/350; 536/23.1; 536/23.5; 536/24.3;		Ronnett, Gabriele V. et al.						
2	435/7.2	435/69.1; 435/7.1; 530/350; 536/23.4		Barak, Lawrence S. et al.						
3	435/7.2	435/69.1; 435/7.1; 530/350; 536/23.4		Barak, Lawrence S. et al.	×					
4	435/194	435/252.3; 435/320.1; 435/6; 536/22.1; 536/23.1; 536/23.2; 536/23.5	,	Chantry, David et al.	×					
5	435/194	435/252.3; 435/320.1; 435/6; 536/22.1; 536/23.1; 536/23.2; 536/23.5	J	Chantry, David et al.	⊠					

RE.CNT 5 THERE ARE 5 CITED REFERENCES AVAILABLE FOR THIS RECORD ALL CITATIONS AVAILABLE IN THE RE FORMAT

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(FILE 'HOME' ENTERED AT 14:31:58 ON 16 AUG 2004)

FILE 'MEDLINE, USPATFULL, CAPLUS' ENTERED AT 14:32:25 ON 16 AUG 2004
L1 0 S DETECT? GPCR AND CONJUGATED ARRESTIN
L2 467 S GPCR AND ARRESTIN
L3 85 S L2 AND GFP
L4 56 S L3 AND PATHWAY
L5 4 S L4 AND PY <2001

=>

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ANSWER 1 OF 4 USPATFULL on STN
L5
       2004:103678 USPATFULL
ΑN
      System for cell-based screening
ΤI
       Dunlay, R. Terry, New Kensington, PA, United States
IN
       Taylor, D. Lansing, Pittsburgh, PA, United States
       Gough, Albert H., Glenshaw, PA, United States
       Giuliano, Kenneth A., Pittsburgh, PA, United States
       Cellomics, Inc., Pittsburgh, PA, United States (U.S. corporation)
PΑ
                               20040427
PΙ
       US 6727071
                          В1
       WO 9838490 19980903
                               19991208 (9)
       US 1999-380259
AΙ
                               19980227
       WO 1998-US3701
       Continuation-in-part of Ser. No. US 1997-810983, filed on 27 Feb 1997,
RLI
       now patented, Pat. No. US 5989835
       US 1997-69329P
                           19971211 (60)
PRAI
                           19971211 (60)
       US 1997-69249P
DТ
       Utility
       GRANTED
FS
LN.CNT 3071
       INCLM: 435/007.210
INCL
       INCLS: 435/007.200; 435/004.000; 435/006.000; 435/007.100; 435/007.500;
              435/288.400; 435/377.000; 435/375.000; 436/010.000; 436/017.000;
              436/063.000; 436/164.000; 436/166.000; 436/172.000; 436/174.000;
              436/517.000; 436/546.000; 382/255.000
NCL
       NCLM:
              435/007.210
              382/255.000; 435/004.000; 435/006.000; 435/007.100; 435/007.200;
       NCLS:
              435/007.500; 435/288.400; 435/375.000; 435/377.000; 436/010.000;
              436/017.000; 436/063.000; 436/164.000; 436/166.000; 436/172.000;
              436/174.000; 436/517.000; 436/546.000
IC
       [7]
       ICM: G01N033-53
       435/7.2; 435/7.21; 435/29; 435/40.5; 435/40.51; 435/288.3; 435/288.4;
EXF
       435/4; 435/7.5; 435/6; 435/7.1; 435/183; 435/375; 435/975; 435/377;
       436/546; 436/172; 436/800; 436/809; 436/10; 436/17; 436/164; 436/166;
       436/174; 436/63; 436/517; 436/56; 356/300; 356/326; 356/328; 382/255;
       382/141; 530/350
CAS INDEXING IS AVAILABLE FOR THIS PATENT.
     ANSWER 2 OF 4 USPATFULL on STN
L5
ΑN
       2000:113721 USPATFULL
       Methods of assaying receptor activity and constructs useful in such
TΙ
       Barak, Lawrence S., Durham, NC, United States
ΙN
       Caron, Marc G., Hillsborough, NC, United States
       Ferguson, Stephen S., London, Canada
       Zhang, Jie, Durham, NC, United States
       Duke University, Durham, NC, United States (U.S. corporation)
PA
                                20000829
PI
       US 6110693
                                19990120 (9)
       US 1999-233530
ΑI
       Continuation of Ser. No. US 1997-869568, filed on 5 Jun 1997, now
RLI
       patented, Pat. No. US 5891646
DΤ
       Utility
FS
       Granted
LN.CNT 1383
       INCLM: 435/007.200
INCL
       INCLS: 536/023.400; 530/350.000; 435/007.100; 435/069.100
              435/007.200
       NCLM:
NCL
              435/007.100; 435/069.100; 530/350.000; 536/023.400
       NCLS:
       [7]
IC
       ICM: G01N033-52
       ICS: C07H021-04; C12N015-12; C07K014-00
       435/7.1; 435/176; 435/183; 435/6; 435/7.2; 435/69.1; 435/320.1;
EXF
       536/23.4; 536/23.5; 530/350
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CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L5 ANSWER 3 OF 4 USPATFULL on STN
AN 1999:43394 USPATFULL

AN 1999:43394 USPATFULL
TI Methods of assaying receptor activity and constructs useful in such methods

IN Barak, Lawrence S., Durham, NC, United States Caron, Marc G., Hillsborough, NC, United States Ferguson, Stephen S., London, Canada Zhang, Jie, Durham, NC, United States

PA Duke University, Durham, NC, United States (U.S. corporation)
PI US 5891646 19990406 <--

AI US 1997-869568 19970605 (8)

DT Utility FS Granted

LN.CNT 1569

INCL INCLM: 435/007.200

INCLS: 536/023.400; 530/350.000; 435/079.100; 435/069.100

NCL NCLM: 435/007.200

NCLS: 435/007.100; 435/069.100; 530/350.000; 536/023.400

IC [6]

ICM: G01N033-52

ICS: C07H021-04; C12N015-12; C07K014-00

EXF 435/71; 435/174; 435/183; 435/6; 435/7.2; 435/69.1; 536/23.4; 536/23.5; 530/350

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L5 ANSWER 4 OF 4 CAPLUS COPYRIGHT 2004 ACS on STN

В1

AN 1998:806798 CAPLUS

DN 130:63343

TI Assaying receptor activity using constructs expressing  $\beta\text{-}$  arrestin conjugates with green fluorescent protein

IN Barak, Lawrence S.; Caron, Marc G.; Ferguson, Stephen S.; Zhang, Jie

PA Duke University, USA

US 2000-631468

SO PCT Int. Appl., 60 pp. CODEN: PIXXD2

DT Patent

LA English

ENN CNIE 2

FAN.CNT 3 PATENT NO.					KIND DATE			AE	APPLICATION NO.						DATE			
ΡΙ		9855635 9855635			A1		1998 2003		WC	) 19	998-1	US11	628		19	9980	604	<
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	US				A		1999	0406	US	3 1	997-	8695	68		1	9970	505	<
	CA	CA 2305810			AA					CA 1998-2305810								
		9877255					1998	1221	JΑ	1 1	998-	7725	5		1:	9980	604	<
	ΑU	AU 759347 EP 1015608			A1 20000705				EP 1998-925260					19980604 <				
	ΕP							EI										
	EP	1015608			В1		2004	0317										
		R: AT,	BE,	CH,	DE,	DK	, ES,	FR,	GB, C	GR,	ΙT,	LI,	LU,	ΝL,	SE,	MC,	PT,	
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	AT	262040			E		2004	AT 1998-925260 EP 2004-6057						19980604				
	ΕP	1441032			A1		2004	0728	E1	- 2	004-	605/	~		2 D	9980	004	
		R: AT,			DE,	DK	, ES,	FR,	GB,	SR,	IT,	ьī,	LU,	ΝL,	SE,	MC,	PT,	
			FI,							_ ^			0.0		^	0000	700	
		20041018							US	5 2	003-	6289	09		2	0030	129	
PRAI		1997-869																
		1998-925					1998											
		1998-US1																
	US	1999-233	530		A1		1999	0120										

20000803

## (FILE 'HOME' ENTERED AT 16:02:42 ON 16 AUG 2004)

FILE 'MEDLINE, BIOSIS, CAPLUS' ENTERED AT 16:03:14 ON 16 AUG 2004 458 S ?ARRESTIN AND GPCR L1L2 34 S L1 AND GFP 7 S L2 AND PY <2000 L3 0 S ARRESATIN AND GFP L4L5234 S ?ARRESTIN AND GREEN FLUORESCENT PROTEIN 51 S L5 AND PY <2000 L6 40 S L6 AND G PROTEIN? L7 34 S L6 AND G PROTEIN-COUPLED RECEPTOR L80 S L8 AND DETECTION 1.9 L10 4 S L8 AND DETECT? 12 S L8 AND ACTIVATION L11=> d 111 1-12 ti py au so abs ANSWER 1 OF 12 MEDLINE on STN Cellular trafficking of G protein-coupled receptor/beta-arrestin endocytic complexes. PΥ 1999 Zhang J; Barak L S; Anborgh P H; Laporte S A; Caron M G; Ferguson S S ΑU SO Journal of biological chemistry, (1999 Apr 16) 274 (16) 10999-1006. Journal code: 2985121R. ISSN: 0021-9258. beta-Arrestins are multifunctional proteins identified on the basis of AB their ability to bind and uncouple G proteincoupled receptors (GPCR) from heterotrimeric G proteins. In addition, beta-arrestins play a central role in mediating GPCR endocytosis, a key regulatory step in receptor resensitization. In this study, we visualize the intracellular trafficking of beta-arrestin2 in response to activation of several distinct GPCRs including the beta2-adrenergic receptor (beta2AR), angiotensin II type 1A receptor (AT1AR), dopamine D1A receptor (D1AR), endothelin type A receptor (ETAR), and neurotensin receptor (NTR). Our results reveal that in response to beta2AR activation, beta-arrestin2 translocation to the plasma membrane shares the same pharmacological profile as described for receptor activation and sequestration, consistent with a role for betaarrestin as the agonist-driven switch initiating receptor endocytosis. Whereas redistributed beta-arrestins are confined to the periphery of cells and do not traffic along with activated beta2AR, D1AR, and ETAR in endocytic vesicles, activation of AT1AR and NTR triggers a clear time-dependent redistribution of beta-arrestins to intracellular vesicular compartments where they colocalize with internalized receptors. Activation of a chimeric AT1AR with the beta2AR carboxyl-terminal tail results in a beta-arrestin membrane localization pattern similar to that observed in response to beta2AR activation. In contrast, the corresponding chimeric beta2AR with the AT1AR carboxyl-terminal tail gains the ability to translocate beta-arrestin to intracellular vesicles. These results demonstrate that the cellular trafficking of beta-arrestin proteins is differentially regulated by the activation of distinct GPCRs. Furthermore, they suggest that the carboxyl-tail of the receptors might be involved in determining the stability of receptor/ betaarrestin complexes and cellular distribution of beta-arrestins.

L11 ANSWER 2 OF 12 MEDLINE on STN

TI Real-time visualization of the cellular redistribution of **G**protein-coupled receptor kinase 2 and betaarrestin 2 during homologous desensitization of the substance P
receptor.

PY 1999

AU Barak L S; Warabi K; Feng X; Caron M G; Kwatra M M

SO Journal of biological chemistry, **(1999 Mar 12)** 274 (11) 7565-9. Journal code: 2985121R. ISSN: 0021-9258.

The substance P receptor (SPR) is a G protein-AΒ coupled receptor (GPCR) that plays a key role in pain regulation. The SPR desensitizes in the continued presence of agonist, presumably via mechanisms that implicate G proteincoupled receptor kinases (GRKs) and beta-arrestins. The temporal relationship of these proposed biochemical events has never been established for any GPCR other than rhodopsin beyond the resolution provided by biochemical assays. We investigate the real-time activation and desensitization of the human SPR in live HEK293 cells using green fluorescent protein conjugates of protein kinase C, GRK2, and beta-arrestin 2. The translocation of protein kinase C betaII-green fluorescent protein to and from the plasma membrane in response to substance P indicates that the human SPR becomes activated within seconds of agonist exposure, and the response desensitizes within 30 s. This desensitization process coincides with a redistribution of GRK2 from the cytosol to the plasma membrane, followed by a robust redistribution of beta-arrestin 2 and a profound change in cell morphology that occurs after 1 min of SPR stimulation. These data establish a role for GRKs and beta-arrestins in homologous desensitization of the SPR and provide the first visual and temporal resolution of the sequence of events underlying homologous desensitization of a GPCR in living cells.

L11 ANSWER 3 OF 12 MEDLINE on STN

TI Using green fluorescent protein to understand the mechanisms of G-protein-coupled receptor regulation.

PY 1998

AU Ferguson S S

SO Brazilian journal of medical and biological research = Revista brasileira de pesquisas medicas e biologicas / Sociedade Brasileira de Biofisica ... [et al.], (1998 Nov) 31 (11) 1471-7. Ref: 50
Journal code: 8112917. ISSN: 0100-879X.

G protein-coupled receptor (GPCR) AΒ activation is followed rapidly by adaptive changes that serve to diminish the responsiveness of a cell to further stimulation. This process, termed desensitization, is the consequence of receptor phosphorylation, arrestin binding, sequestration and down-regulation. GPCR phosphorylation is initiated within seconds to minutes of receptor activation and is mediated by both second messenger-dependent protein kinases and receptor-specific G protein-coupled receptor kinases (GRKs). Desensitization in response to GRK-mediated phosphorylation involves the binding of arrestin proteins that serve to sterically uncouple the receptor from its G protein. GPCR sequestration, the endocytosis of receptors to endosomes, not only contributes to the temporal desensitization of GPCRs, but plays a critical role in GPCR resensitization. GPCR down-regulation, a loss of the total cellular complement of receptors, is the consequence of both increased lysosomal degradation and decreased mRNA synthesis of GPCRs. While each of these agonist-mediated desensitization processes are initiated within a temporally dissociable time frame, recent data suggest that they are intimately related to one another. The use of green fluorescent protein from the jellyfish Aqueora victoria as an epitope tag with intrinsic fluorescence has facilitated our understanding of the relative relationship between GRK phosphorylation,

arrestin binding, receptor sequestration and down-regulation.

L11 ANSWER 4 OF 12 MEDLINE on STN

TI A beta-arrestin/green fluorescent

protein biosensor for detecting G proteincoupled receptor activation.

- PY 1997
- AU Barak L S; Ferguson S S; Zhang J; Caron M G
- SO Journal of biological chemistry, (1997 Oct 31) 272 (44) 27497-500.

Journal code: 2985121R. ISSN: 0021-9258.

AB G protein-coupled receptors (GPCR)

represent the single most important drug targets for medical therapy, and information from genome sequencing and genomic data bases has substantially accelerated their discovery. The lack of a systematic approach either to identify the function of a new GPCR or to associate it with a cognate ligand has added to the growing number of orphan receptors. In this work we provide a novel approach to this problem using a beta-arrestin2/green fluorescent protein

conjugate (betaarr2-GFP). It provides a real-time and single cell based assay to monitor GPCR  ${\bf activation}$  and  ${\tt GPCR-G}$ 

protein-coupled receptor kinase or GPCR-

arrestin interactions. Confocal microscopy demonstrates the translocation of betaarr2-GFP to more than 15 different ligand-activated GPCRs. These data clearly support the common hypothesis that the beta-arrestin binding of an activated receptor is a convergent step of GPCR signaling, increase by 5-fold the number of GPCRs known to interact with beta-arrestins, demonstrate that the cytosol is the predominant reservoir of biologically active beta-arrestins, and provide the first direct demonstration of the critical importance of **G** 

protein-coupled receptor kinase

phosphorylation to the biological regulation of beta-arrestin activity and GPCR signal transduction in living cells. The use of betaarr2-GFP as a biosensor to recognize the activation of pharmacologically distinct GPCRs should accelerate the identification of orphan receptors and permit the optical study of their signal transduction biology intractable to ordinary biochemical methods.

- L11 ANSWER 5 OF 12 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN
- TI Activation and desensitisation of the thyrotropin-releasing hormone receptor visualised by monitoring cellular redistribution of a beta-arrestin-l-green fluorescent protein fusion protein.
- PY 1999
- AU Groarke, Alex [Reprint author]; Milligan, Graeme [Reprint author]
- Biochemical Society Transactions, (1999) Vol. 27, No. 3, pp. A118. print. Meeting Info.: 668th Meeting of the Biochemical Society. Glasgow, Scotland, UK. April 7-9, 1999. Biochemical Society. CODEN: BCSTB5. ISSN: 0300-5127.
- L11 ANSWER 6 OF 12 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN
- TI Real-time visualization of the cellular redistribution of G
  protein-coupled receptor kinase 2 and betaarrestin 2 during homologous desensitization of the substance P
  receptor.
- PY **1999**
- AU Barak, Larry S.; Warabi, Kengo; Feng, Xiao; Caron, Marc G.; Kwatra, Madan M. [Reprint author]
- SO Journal of Biological Chemistry, (March 12, 1999) Vol. 274, No. 11, pp. 7565-7569. print. CODEN: JBCHA3. ISSN: 0021-9258.
- AB The substance P receptor (SPR) is a **G protein**coupled receptor (GPCR) that plays a key role in pain
  regulation. The SPR desensitizes in the continued presence of agonist,
  presumably via mechanisms that implicate **G protein**coupled receptor kinases (GRKs) and beta-arrestins. The
  temporal relationship of these proposed biochemical events has never been
  established for any GPCR other than rhodopsin beyond the resolution

provided by biochemical assays. We investigate the real-time activation and desensitization of the human SPR in live HEK293 cells using green fluorescent protein conjugates of protein kinase C, GRK2, and beta-arrestin 2. translocation of protein kinase C betaII-green fluorescent protein to and from the plasma membrane in response to substance P indicates that the human SPR becomes activated within seconds of agonist exposure, and the response desensitizes within 30 s. This desensitization process coincides with a redistribution of GRK2 from the cytosol to the plasma membrane, followed by a robust redistribution of beta-arrestin 2 and a profound change in cell morphology that occurs after 1 min of SPR stimulation. These data establish a role for GRKs and beta-arrestins in homologous desensitization of the SPR and provide the first visual and temporal resolution of the sequence of events underlying homologous desensitization of a GPCR in living cells.

- L11 ANSWER 7 OF 12 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN
- TI Using green fluorescent protein to understand the mechanisms of G-protein-coupled receptor regulation.
- PY 1998
- AU Ferguson, S. S. G. [Reprint author]
- SO Brazilian Journal of Medical and Biological Research, (Nov., 1998) Vol. 31, No. 11, pp. 1471-1477. print. CODEN: BJMRDK. ISSN: 0100-879X.
- AB G protein-coupled receptor (GPCR)

activation is followed rapidly by adaptive changes that serve to diminish the responsiveness of a cell to further stimulation. This process, termed desensitization, is the consequence of receptor phosphorylation, arrestin binding, sequestration and down-regulation. GPCR phosphorylation is initiated within seconds to minutes of receptor activation and is mediated by both second messenger-dependent protein kinases and receptor-specific G protein-coupled receptor kinases (GRKs).

Desensitization in response to GRK-mediated phosphorylation involves the binding of arrestin proteins that serve to sterically uncouple the receptor from its G protein. GPCR sequestration, the endocytosis of receptors to endosomes, not only contributes to the temporal desensitization of GPCRs, but plays a critical role in GPCR resensitization. GPCR down-regulation, a loss of the total cellular complement of receptors, is the consequence of both increased lysosomal degradation and decreased mRNA synthesis of GPCRs. While each of these agonist-mediated desensitization processes are initiated within a temporally dissociable time frame, recent data suggest that they are intimately related to one another. The use of green

fluorescent protein from the jellyfish Aqueora victoria as an epitope tag with intrinsic fluorescence has facilitated our understanding of the relative relationship between GRK phosphorylation, arrestin binding, receptor sequestration and down-regulation.

- L11 ANSWER 8 OF 12 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN
- TI A beta-arrestin/green fluorescent protein biosensor for detecting G protein-coupled receptor activation.
- PY 1997
- AU Barak, Larry S.; Ferguson, Stephen S. G.; Zhang, Jie; Caron, Marc G. [Reprint author]
- SO Journal of Biological Chemistry, (Oct. 31, 1997) Vol. 272, No. 44, pp. 27497-27500. print. CODEN: JBCHA3. ISSN: 0021-9258.
- AB **G protein-coupled receptors** (GPCR) represent the single most important drug targets for medical therapy, and information from genome sequencing and genomic data bases has

substantially accelerated their discovery. The lack of a systematic approach either to identify the function of a new GPCR or to associate it with a cognate ligand has added to the growing number of orphan receptors. In this work we provide a novel approach to this problem using a beta-arrestin2/green fluorescent protein conjugate (betaarr2-GFP). It provides a real-time and single cell based assay to monitor GPCR activation and GPCR-G protein-coupled receptor kinase or GPCRarrestin interactions. Confocal microscopy demonstrates the translocation of betaarr2-GFP to more than 15 different ligand-activated GPCRs. These data clearly support the common hypothesis that the betaarrestin binding of an activated receptor is a convergent step of GPCR signaling, increase by 5-fold the number of GPCRs known to interact with beta-arrestins, demonstrate that the cytosol is the predominant reservoir of biologically active beta-arrestins, and provide the first direct demonstration of the critical importance of  ${\bf G}$ protein-coupled receptor kinase phosphorylation to the biological regulation of beta-arrestin activity and GPCR signal transduction in living cells. The use of betaarr2-GFP as a biosensor to recognize the activation of pharmacologically distinct GPCRs should accelerate the identification of orphan receptors and permit the optical study of their signal transduction

- L11 ANSWER 9 OF 12 CAPLUS COPYRIGHT 2004 ACS on STN
- TI Real-time visualization of the cellular redistribution of  ${\bf G}$  protein-coupled receptor kinase 2 and  $\beta-$  arrestin 2 during homologous desensitization of the substance P receptor
- PY 1999
- AU Barak, Larry S.; Warabi, Kengo; Feng, Xiao; Caron, Marc G.; Kwatra, Madan M.
- SO Journal of Biological Chemistry (1999), 274(11), 7565-7569 CODEN: JBCHA3; ISSN: 0021-9258

biology intractable to ordinary biochemical methods.

- AΒ The substance P receptor (SPR) is a G proteincoupled receptor (GPCR) that plays a key role in pain regulation. The SPR desensitizes in the continued presence of agonist, presumably via mechanisms that implicate G proteincoupled receptor kinases (GRKs) and  $\beta$ -arrestins. The temporal relationship of these proposed biochem. events has never been established for any GPCR other than rhodopsin beyond the resolution provided by biochem. assays. The authors investigate the real-time activation and desensitization of the human SPR in live HEK293 cells using green fluorescent protein conjugates of protein kinase C, GRK2, and  $\beta$ - arrestin 2. The translocation of protein kinase C  $\beta \text{II-}$  green fluorescent protein to and from the plasma membrane in response to substance P indicates that the human SPR becomes activated within seconds of agonist exposure, and the response desensitizes within 30 s. This desensitization process coincides with a redistribution of GRK2 from the cytosol to the plasma membrane, followed by a robust redistribution of  $\beta$ - arrestin 2 and a profound change in cell morphol. that occurs after 1 min of SPR stimulation. These data establish a role for GRKs and  $\beta\text{-arrestins}$  in homologous desensitization of the SPR and provide the first visual and temporal resolution of the sequence of events underlying homologous desensitization of a GPCR in living cells.
- L11 ANSWER 10 OF 12 CAPLUS COPYRIGHT 2004 ACS on STN
- TI Using green fluorescent protein to understand the mechanisms of G-protein-coupled receptor regulation
- PY 1998
- AU Ferguson, S. S. G.

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Brazilian Journal of Medical and Biological Research (1998),
     31(11), 1471-1477
     CODEN: BJMRDK; ISSN: 0100-879X
     A review, with 50 refs. G protein-coupled
AΒ
     receptor (GPCR) activation is followed rapidly by
     adaptive changes that serve to diminish the responsiveness of a cell to
     further stimulation. This process, termed desensitization, is the
     consequence of receptor phosphorylation, arrestin binding,
     sequestration and down-regulation. GPCR phosphorylation is initiated
     within seconds to minutes of receptor activation and is mediated
     by both second messenger-dependent protein kinases and receptor-specific
     G protein-coupled receptor kinases
     (GRKs). Densensitization in response to GRK-mediated phosphorylation
     involves the binding of arrestin proteins that serve to
     sterically uncouple the receptor from its G protein. GPCR sequestration,
     the endocytosis of receptors to endosomes, not only contributes to the
     temporal desensitization of GPCRs, but plays a critical role in GPCR
     resensitization. GPCR down-regulation, a loss of the total cellular
     complement of receptors, is the consequence of both increased lysosomal
     degradation and decreased mRNA synthesis of GPCRs. While each of these
     agonist-mediated desensitization processes are initiated within a
     temporally dissociable time frame, recent data suggest that they are
     intimately related to one another. The use of green
     fluorescent protein from the jellyfish Aqueora victoria
     as an epitope tag with intrinsic fluorescence has facilitated our
     understanding of the relative relationship between GRK phosphorylation,
     arrestin binding, receptor sequestration and down-regulation.
L11 ANSWER 11 OF 12 CAPLUS COPYRIGHT 2004 ACS on STN
     Assaying receptor activity using constructs expressing \beta-
     arrestin conjugates with green fluorescent
     protein
PΥ
     1998
     2003
     1999
     1998
     1998
     2003
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     2004
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     2004
     Barak, Lawrence S.; Caron, Marc G.; Ferguson, Stephen S.; Zhang, Jie
ΙN
SO
     PCT Int. Appl., 60 pp.
     CODEN: PIXXD2
     Described are methods of detecting G-protein
AB
     coupled receptor (GPCR) activity in vivo and in vitro;
     methods of assaying GPCR activity; and methods of screening for GPCR
     ligands, G protein-coupled receptor
     kinase (GRK) activity, and compds. that interact with components of the
     GPCR regulatory process. The assays are based on the observation that
     \beta- arrestin redistribution from the cytosol to the plasma
     membrane occurs in response to agonist activation of GPCRs,
     demonstrating a common role for \beta- arrestin in
     agonist-mediated signal transduction termination. Thus, translocation of
     conjugates of a \beta- arrestin protein with a detectable mol.
     indicates activation of the GPCR pathway. A fusion protein
     consisting of a \beta- arrestin mol. (\beta-arrestin2)
     conjugated to a green fluorescent protein
     (S65T variant) at its C-terminus is expressed in cells and is biol.
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active. This conjugates allows convenient methods of assaying agonist

stimulation of GPCRS in vivo and in vitro in real time.

- L11 ANSWER 12 OF 12 CAPLUS COPYRIGHT 2004 ACS on STN
- TI A  $\beta$  arrestin/green fluorescent protein biosensor for detecting G protein-coupled receptor activation
- PY 1997
- AU Barak, Larry S.; Ferguson, Stephen S. G.; Zhang, Jie; Caron, Marc G.
- SO Journal of Biological Chemistry (1997), 272(44), 27497-27500 CODEN: JBCHA3; ISSN: 0021-9258
- AB G protein-coupled receptors (GPCR)

represent the single most important drug targets for medical therapy, and information from genome sequencing and genomic data bases has substantially accelerated their discovery. The lack of a systematic approach either to identify the function of a new GPCR or to associate it with a cognate ligand has added to the growing number of orphan receptors. In this work we provide a novel approach to this problem using a  $\beta$ -arrestin2/ green fluorescent protein

conjugate ( $\beta$ arr2-GFP). It provides a real-time and single cell based assay to monitor GPCR activation and GPCR-G

protein-coupled receptor kinase or GPCR-

arrestin interactions. Confocal microscopy demonstrates the translocation of  $\beta arr2\text{-}GFP$  to more than 15 different ligand-activated GPCRs. These data clearly support the common hypothesis that the  $\beta\text{-}$  arrestin binding of an activated receptor is a convergent step of GPCR signaling, increase by 5-fold the number of GPCRs known to interact with  $\beta\text{-}$  arrestins, demonstrate that the cytosol is the predominant reservoir of biol. active  $\beta\text{-}$  arrestins, and provide the first direct demonstration of the critical importance of G protein-coupled receptor kinase phosphorylation to the biol. regulation of  $\beta\text{-}$  arrestin activity and GPCR signal transduction in living cells. The use of  $\beta$  arr2-GFP as a biosensor to recognize the activation of pharmacol. distinct GPCRs should accelerate the identification of orphan receptors and permit the optical study of their signal transduction biol. intractable to ordinary biochem. methods.